

U.S. Micro-Solutions, Inc. * 302 Unity Plaza * Latrobe, PA 15650 Phone: (724) 853-4047 Fax: (724) 853-4049 AIHA-LAP, LLC EMLAP # 103009 www.usmslab.com





Customer Name: AGX Inc. Sample Date: August 18, 2020
Customer Address: 207 Pine Creek Road Date Received: August 19, 2020
Wexford, PA 15090 Date of Report: August 24, 2020

Customer Phone: (724) 934-4249 Fax: (724) 934-5677
PO Number: Attention: Amber Brancolini

Project Name/Number: Connoquenessing Elementary School

Customer sample numbers below are uniquely identified by prefixing Laboratory # 86769-20														
Gustomer cump			Trap Analysis Analytical M	;	-		AllergencoD							
Total Volume (L)	<u> </u>		75	ctiloa.	75				ı		75			
Sample Number		CONNO-01				C	ONNO-02			C	ONNO-03			
Location:			Room 20				Room 1				Room 6			
Particle ID	Raw ct.	AS	Spores/m³	%	Raw ct.	AS	Spores/m³	%	Raw ct.	AS	Spores/m³	%		
Alternaria														
Ascospores	2	13	26	5%					2	13	26	4%		
Aspergillus/Penicillium-like	2	13	26	5%										
Basidiospores	31	13	403	84%	45	13	585	90%	46	13	598	96%		
Bipolaris/Drechslera	0.			0.70	.0			0070	.0	.0		0070		
Cercospora														
Chaetomium														
Cladosporium	2	13	26	5%	2	13	26	4%						
Curvularia		10	20	370		10	20	470						
Epicoccum														
Helicomyces														
Nigrospora														
Oidium														
					4	40	42	20/						
Pithomyces/Ulocladium					1	13	13	2%						
Polythrincium														
Rusts														
Smuts/ Myxomycetes														
Stachybotrys														
Torula														
Trichoderma Unidentified dematiaceous conidia					2	13	26	4%						
Unidentified hyaline conidia					2	10	20	470						
,														
Total Mold (Spores/m³ of air)	37		481		50		650		48		624			
		40	40	1	_	40	40			40	40			
Pollen	0	13	< 13		0	13	< 13		0	13	< 13			
Hyphal Fragments														
Insect Fragments														
Plant Fragments														
Skin Cell Fragments			1		1					1				
Debris Debris			1		1					1				
Analyst Initials			JM				JM				JM			
Date Analyzed		-	08/21/20				08/21/20	-			08/21/20			
Cassette Serial # / Exp Date:		322	0371 10/2020			322	0375 10/2020	3220375 10/2020			3220359 10/2020			

Entire trace analyzed. Results relate only to the samples tested. Results are reported as calculated. For biological data, the first and/or second digit should be considered significant. Total percentage may not equal 100% due to rounding. Percentages reported as 0% are greater than 0 and less than 0.5%. The Aspergillus/PenicIlium-like category cannot be differentiated by non-viable sampling methods.

AS=Analytical Sensitivity (spores/m³); Blank Lines = None Detected

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Technical Manager: Flown Damko
Sharon Danko, AS, MLT (ASCP)



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Customer sampl	_		_	-		hy prof	fiving Labora	tory#		86769	a_20	
Customer Sampi			Trap Analysis Analytical M	i	-	MIC 01	AllergencoD	itory #		3070	<i>,</i> - <u>2</u> 0	
Total Volume (L)			75	etilou.	I	WIIO U	75		l		75	
Sample Number		C	ONNO-04			C	CONNO-05			C	ONNO-06	
Location:		Fac	ulty Lounge			1	Room 17				Room 14	
Particle ID	Raw ct.	AS	Spores/m³	%	Raw ct.	AS	Spores/m³	%	Raw ct.	AS	Spores/m³	%
Alternaria												
Ascospores					5	13	65	20%	4	13	52	12%
Aspergillus/Penicillium-like												
Basidiospores	83	13	1,079	97%	12	13	156	48%	25	13	325	74%
Bipolaris/Drechslera												
Cercospora												
Chaetomium												
Cladosporium	3	13	39	3%	4	13	52	16%	5	13	65	15%
Curvularia												
Epicoccum					1	13	13	4%				
Helicomyces					·			1,0				
Nigrospora												
Oidium												
Pithomyces/Ulocladium												
Polythrincium												
Rusts												
					2	40	00	00/				
Smuts/ Myxomycetes					2	13	26	8%				
Stachybotrys												
Torula Trichoderma												
Unidentified dematiaceous conidia					1	13	13	4%				
Unidentified hyaline conidia								.,,				
Total Mold (Spores/m³ of air)	86		1,118		25		325		34		442	
(C) C C C C C C C C C C C C C C C C C C			,	_								
Pollen	0	13	< 13		0	13	< 13		0	13	< 13	
Hyphal Fragments												
Insect Fragments												
Plant Fragments												
Skin Cell Fragments			1		1					1		
Debris			1		3***					2		
Analyst Initials			JM				JM				JM	
Date Analyzed			08/21/20				08/21/20				08/21/20	
Cassette Serial # / Exp Date: Entire trace analyzed Results relate only			0366 10/2020				0369 10/2020			322	0374 10/2020	

Entire trace analyzed. Results relate only to the samples tested. Results are reported as calculated. For biological data, the first and/or second digit should be considered significant. Total percentage may not equal 100% due to rounding. Percentages reported as 0% are greater than 0 and less than 0.5%. The Aspergillus/Penicllium-like category cannot be differentiated by non-viable sampling methods.

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***A debris rating of 3 or greater indicates that the accuracy of the analysis is likely affected.

Technical Manager: Flavon Damko
Sharon Danko, AS, MLT (ASCP)



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Customer sample numbers below are uniquely identified by prefixing Laboratory # 86769-20												
	Airborne	Spore	Trap Analysis Analytical M	;	- AllergencoD							
Total Volume (L)			75	ctilou.	ı	14110 01	75		ı			
Sample Number		(CONNO-07			C	ONNO-08					
Location:			Library				Outside					
Particle ID	Raw ct.	AS	Spores/m³	%	Raw ct.	AS	Spores/m³	%	Raw ct.	AS	Spores/m³	%
Alternaria												
Ascospores	1	13	13	7%	120	13	1,560	5%				
Aspergillus/Penicillium-like				.,,	12	13	156	0%				
Basidiospores	7	13	91	47%	105	266	27,930	88%				
Bipolaris/Drechslera	- '	13	91	41 70	105	200	21,930	00 /0				
					4	40	40	00/				
Cercospora					1	13	13	0%				
Chaetomium				400/				=0/				
Cladosporium	6	13	78	40%	171	13	2,223	7%				
Curvularia												
Epicoccum												
Helicomyces												
Nigrospora												
Oidium												
Pithomyces/Ulocladium												
Polythrincium												
Rusts					1	13	13	0%				
Smuts/ Myxomycetes												
Stachybotrys												
Torula												
Trichoderma												
Unidentified dematiaceous conidia	1	13	13	7%								
Unidentified hyaline conidia												
Total Mold												
(Spores/m³ of air)	(Spores/m³ of air) 15 195 410 31,895											
Pollen	0	13	< 13		3	13	39					
Hyphal Fragments												
Insect Fragments												
Plant Fragments												
Skin Call Fragment			4									
Skin Cell Fragments Debris			1		0							
Analyst Initials			JM				JM					
Date Analyzed			08/21/20				08/21/20					
Cassette Serial # / Exp Date:		322	20364 10/2020			321	9485 10/2020					

Entire trace analyzed. Results relate only to the samples tested. Results are reported as calculated. For biological data, the first and/or second digit should be considered significant. Total percentage may not equal 100% due to rounding. Percentages reported as 0% are greater than 0 and less than 0.5%. The Aspergillus/PenicIlium-like category cannot be differentiated by non-viable sampling methods.

AS=Analytical Sensitivity (spores/m³); Blank Lines = None Detected

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Technical Manager: Sharon Danko, AS, MLT (ASCP)

SPORE TRAP INTERPRETATION TIPS

Contains opinions and interpretations

Currently there are no numeric standards for indoor airborne or surface microbial contamination. Suggested guidelines are constantly being reviewed and updated as more information is collected.

Some common denominators should be considered when interpreting results:

- 1. Comparison of indoor/outdoor concentration ratios.
- 2. Complaint vs. non-complaint areas or affected vs. non-affected areas.
- 3. Consider air exchange rates and activity levels in a building structure, weather, and season of the year.
- 4. Rank order assessment and concentration (e.g. Spores/m3 of air) of the fungi.
- 5. Predominant fungal genera: Are there water indicator microorganisms present, such as but not limited to: Chaetomium, Stachybotrys, Rhodotorula, Trichoderma, and Scopulariopsis.
- 6. Generally fungal counts indoors should be lower than outdoor counts and the types of fungi found indoors should be similar to outdoors.
- 7. There is always a potential bias from infiltration of outdoor air, poor housekeeping, excessive indoor relative humidity, or potential contamination sources (e.g. water intrusion through a basement wall) that may negatively influence post remedial verification (PRV) or clearancelevels.
- 8. The investigator should look for various patterns among the indoor types of molds detected:
 - a. Increased levels of primary (1st) colonizers in damp or moisture intrusion areas of homes or commercial buildings: **Aspergillus/Penicillium** or **Cladosporium** are usually noted.
 - b. Chaetomium or Stachybotrys are tertiary (3rd) colonizers of indoor materials and are usually associated with chronic long-standing water/moisture issues in a building.
 - c. The presence of **hyphal fragments** or **fruiting structures** noted on spore trap samples usually indicates amplification (growth) of fungi on building substrates.
 - d. **Ascospores** and **basidiospores** noted on indoor spore trap samples most often represent the entrance of inadequately filtered outdoor air. During inclement weather, remember to note time, temperature, and season. Most indoor materials will not support the growth of these fungi.
- 9. When unidentified **hyaline** (clear) or **dematiaceous** (dark-pigmented) conidia are noted on a spore trap sample, it indicates that no particular fungus can be identified. These fungal conidia may represent such yeast-like fungi as *Aureobasidium*, *Sporidiobolus*, unidentifiable *Acremonium* species, Basidiomycetes (basidiospores), and Ascomycetes (ascospores).
- 10. Keep in mind when interpreting spore trap sample reports, that indoor levels may be higher than corresponding outdoor levels (winter time in the northern U.S.) with a predominance of *Aspergillus/Penicillium* or *Cladosporium* conidia with no significant amplification of any molds.

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SPORE TRAP GUIDELINES FOR INDOOR MICROBIAL CONTAMINATION

	DEBRIS RATING for SPORE TRAP ANALYSIS (Air-O-Cell, Micro 5, Allergenco D, Cyclex	
DEBRIS RATING	CONDITIONS FOR REPORTING DEBRIS RATING	SIGNIFICANCE
0	A visible trace, including particulates and debris, is not observed.	Indicates the sample is a blank, the area is exceptionally clean, or improper sampling occurred.
1	Debris is present and <10% of the average viewing field is obscured.	Minimal amount of debris is observed.
2	Debris is present and 10% to <40% of the average viewing field is obscured.	Low amount of debris is observed, counts may be affecte
3*	Debris is present and 40% to 75% of the average viewing field is obscured.	Moderate amount of debris is observed, counts of conidia/hyphal fragments may be underestimated.
4*	Debris is present and >75% of the average viewing field is obscured.	High amount of debris is observed, counts are estimated
5* See Relative Abundance chart below	Excessive debris is present.	Periphery of trace analyzed. Relative amounts of conidia/hyphal fragments noted. Suggest recollection
6	Slide completely obscured by excessive debris.	Unable to analyze. Recollect sample.

^{*}A rating of 3 or greater indicates that the accuracy of the analysis is likely affected.

RELATIVE ABUNDANCE of OBSERVED CONIDIA & HYPHAL FRAGMENTS						
RATING	Relative Amounts of Observed Fungal Structures per high power field (600 X)					
Rare	0-1					
Few	2 to 5					
Moderate	6 to 10					
Many	11 to 100					
Numerous	>100					

SKIN CELL ANALYSIS					
SKIN CELL RATING	Relative Amounts of Observed Skin Cells per high power field (600 X)				
0	No skin cells present				
1	0-1				
2	2 to 5				
3	6 to 10				
4	11 to 15				
5	≥16				

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DCR 20-079

Effective 06-04-20

End of Report





U.S. Micro-Solutions, Inc. 302 Unity Plaza Latrobe, PA 15650 PH: 724-853-4047 FAX: 724-853-4049





supplies@usmslab.com

LABORATORY TEST REQUEST - CHAIN OF CUSTODY

			erm v		-		
Customer Name:	AGX Inc.			Phone #: 724-934-4249	FAX#: 72	4-934-567	7
Address:	207 Pine Cre	ek Road		City: Wexford	Zip: 15	090	
Attention To:	Amber Brand	colini		E-Mail: abrancolini@agxinc.com			
Sample Obtained	By: Amber	Brancolini		Results: FAX	PO#	Proposal	#
Project Name/Nu	ımber: Conno	evenessin	? Elemer	Hary School	а		
Turn-Around-Tim (Spore Trap & DN		Standard (48-	-72 hr) N	ext Day (24 hr, M-F) Same Day (6 hr, M	1-F) 3-H	our (M-F)	Saturday
Comments:							
Sample #	Sample Date / Time	Sample Code	Analysis Code	Sample Location & Des	cription		Sample Volume/Area
Conno - 01	8-18-20	ST	SPT	Room 20	V		75 L
conno-02				Room 1		*	
(onno-03				Room 6			
(onn 0-64				Faculty Res Lounge			
conno-05				Room 17			
Conno-06				Room 14.			
Conno-07				Library			
Conno-08	1	4	1	outside			1
Relinquished By	(Customer MUST sig	auls	Brion	restii		ate & Time - 18-20 /	4:35 PM
Received By – La	ab Use Only		Mar	Date & Time		# 0 (0920
Rev. 12-14-17		A	1	Tikit in all			

Sample Code					
A Air Plate					
B Bulk					
Spore Trap					
Swab					
Water					
Таре					
Other					

Analysis Code							
DME	Direct Microscopic Exam	НРС	Heterotrophic Plate Count				
SPT	Spore Trap	MYC	Mycobacteria Culture				
FUNG	Fungal Culture – Counts w/ ID of top 3 organisms	STA	Staphylococcus / MRSA Culture				
BACT	Bacterial Culture – Counts w/ ID of top 3 organisms	DUO	Duodenoscope Culture				
SSQT	Sewage Screen (quant) – Counts w/ Identification E. coli, coliforms, enterococci (fecal streptococci)	HCU	Heater/Cooler Water Culture includes mycobacteria, HPC, coliforms, & P. aeruginosa				
SSQL	Sewage Screen (qualitative) – Identification of E. coli, coliforms, enterococci (fecal streptococci)	PSA	Pseudomonas aeruginosa Culture				
COL	Colilert - Presence/absence of E. coli, coliforms	IDS	Species Identification by MALDI-TOF				

^{*}All samples received after 1:00 p.m. Monday-Friday will be considered received the NEXT business day. Same Day and Next Day samples received on Saturday will be reported on Monday and Tuesday, respectively.